TERPENOIDS OF HELIANTHUS NUTTALLII

Esther F. Lee, Jonathan Gershenzon, and Tom J. Mabry $% \mathcal{F}_{\mathcal{F}}$

Department of Botany, University of Texas at Austin, Austin, TX 78713

In conjunction with our continuing study of the terpenoid chemistry of Helianthus (Asteraceae), we report here the results of our investigation of Helianthus nuttallii T. & G. ssp. nuttallii. A large number of sesquiterpene lactones and diterpenes have been isolated previously from species of Helianthus (1-6, and references cited therein). H. nuttallii is a diploid perennial native to southern Canada and the Rocky Mountain region of the United States (7) and is classified in section Divaricati series Corona-Solis in the most recent taxonomic revision of the genus (8). Based on morphological data and the results of crossing studies, H. nuttallii is considered to be most closely related to Helianthus grosseserratus Martens and Helianthus giganteus L. (7,9,10). Systematically, these three species form a taxonomic complex that, according to Heiser (7), could be treated as a highly variable single species, although he has maintained them as separate entities.

Three diterpene carboxylic acids and two sesquiterpene lactones were isolated from a CH₂Cl₂ extract of the aerial parts of H. nuttalli ssp. nuttallii: 15a-OH-entkaur-16(17)-en-19-oic acid (grandifloric acid), 1 (2,11); 17-OH-ent-isokaur-15(16)-en-19-oic acid, 3(12); 7α-OHent-trachvloban-19-oic acid (ciliaric acid), 5(13); angeloyloxyatripliciolide, 6(14,15); and 1α -OH-2 β -OH-dihydropinnatifidin, 9(16). Comparison of our results with those obtained earlier from H. grosseserratus (16) supports the proposed close relationships between H. nuttallii and H. grosseseratus. Compounds 1, 3, 5, 7, 8, 9, 10, and 11 were reported from H. grosseserratus by Herz and co-workers (16); we also isolated five of these, namely 1, 3, 5, 7, and 9, from this same species (17). Both taxa, H. nuttallii and H. grosseserratus, contain

furanoheliangolides, 12,8-cis-lactonized eudesmanolides, and the same three diterpene carboxylic acids. The unusual 12,8-cis-lactonized eudesmanolide 9 provides strong chemical evidence for the proposed close relationship of these species. Compounds 9, 10, and 11 are the first eudesmanolides and the first cislactonized sesquiterpene lactones isolated from this genus.

In contrast, our chemical studies do not support a close relationship between *H. nuttallii* and *H. giganteus*. *H. giganteus* yielded two 1,2-secogermacranolides (18). An earlier examination of the latter species (19) reported only the sesquiterpene germacrene D from the aerial parts and kaurane diterpenes from the roots.

EXPERIMENTAL

PLANT MATERIAL.—Leaves of *H. nuttallii* ssp. nuttallii were collected at the U. S. Department of Agriculture research station at Bushland, Texas, on August 11, 1979, and August 2, 1980, from plants growing from rootstocks transplanted from Colorado (8.5 miles north of Louviers on Hwy 85, Douglas Co., August 22, 1977, C.E. Rogers and T.E. Thompson #734) and Utah (junction of interstate Hwy 15 and state Hwy 115 near Payson, Utah Co., August 24, 1977, C.E. Rogers and T.E. Thompson #750). Voucher specimens, J.G. #84 and #85, respectively, are deposited in the Herbarium of the University of Texas at Austin.

EXTRACTION AND FRACTIONATION.—TIc profiles of leaf wash extracts of the two collections were very similar. Therefore, all of the plant material (1.9 kg) was combined and washed with CH_2Cl_2 for 5 min at room temperature. Intact rather than ground leaves were extracted, since in many species of *Helianthus* sesquiterpene lactones are localized in surface glands (20) and a rapid surface wash has been shown to give a greater absolute yield of sesquiterpene lactones and reduced amounts of other plant constituents than does an extraction of ground material. Qualitative differences in terpenoid constituents are not found between leaf wash and ground whole-leaf extracts. The wash was concentrated in vacuo and was pre-



pared for column chromatography by standard procedures (21) to yield 20.7 g of yellow syrup. The syrup was chromatographed over a silica gel column (1 kg) packed in CH_2Cl_2 . The column was eluted with a CH_2Cl_2 -iPrOH gradient. Fifty fractions of one liter each were collected.

Crystals formed in fractions 17-20 (1.5% iso-PrOH), which upon recrystallization from isopropyl ether-EtOAc, gave 85 mg of 6, mp 134° {lit 132-134° (14,15)}. In fractions 22 and 23 (2% iPrOH) a precipitate formed which, after washing in cold MeOH, gave 10 mg of 5. A precipitate from fractions 27 and 28 (5% iPrOH) was washed in cold MeOH to give 7 mg of a 1:1 mixture of 1 and 3 (5% iPrOH). Because the quantity of this material was limited, an additional 30 mg of the same mixture (identity established by nmr and tlc) was obtained from *H. grosseserratus* (17). Methylation with CH₂N₂ and separation of the reaction products by preparative tlc (silica gel, 1 mm, CHCl₃-MeOH, 15:1) gave 7 mg of 2 and 7 mg of 4. Crystals of 9 (385 mg), mp 218° {lit 222-224° (16)}, were obtained from fractions 30 and 31 (5% iPrOH) after recrystallization from hot EtOAc. All compounds were identified by comparison of their spectral data with those in the literature {1(2, 11), 2(2), 4(12), 5(13), 6(14, 15), and 9(16)]. Compounds 2, 4, and **9** were also identified by co-tlc with authentic samples isolated from other species of *Helianthus* (2,3,17).

ACKNOWLEDGMENTS

We thank Drs. C. Rogers, G. Seiler, and T. Thompson for collecting and cultivating the plants, J. Hudson and M. Leidig for ms measurements, B.A. Shoulders for nmr measurements, and the National Institutes of Health (Grant HDO-4488) and the Robert A. Welch Foundation (Grant F-130) for financial support.

LITERATURE CITED

- N. Ohno and T.J. Mabry, *Phytochemistry*, 18, 1003 (1979).
- N. Ohno, T.J. Mabry, V. Zabel, and W.H. Watson, *Phytochemistry*, 18, 1687 (1979).
- N. Ohno and T.J. Mabry, *Phytochemistry*, 19, 609 (1980).
- N. Ohno, J. Gershenzon, P. Neuman, and T.J. Mabry, *Phytochemistry*, **20**, 2393 (1980).
- K. Watanabe, N. Ohno, H. Yoshioka, J. Gershenzon, and T.J. Mabry, *Phytochemistry*. 21, 709 (1982).
- 6. J. Gershenzon and T.J. Mabry, *Phytochem-istry*. 23, 1959 (1984).
- C.B. Heiser, D.M. Smith, S.B. Clevenger, and W.C. Martin, Jr., Mem. Torr. Bot. Club. 22, 1 (1969).
- E.E. Schilling and C.B. Heiser, Taxon, 30, 393 (1981).
- 9. C.B. Heiser, W.C. Martin, and D.M. Smith, *Brittonia*. 14, 137 (1962).

- 10. T.E. Long, Brittonia, 18, 64 (1966).
- F. Piozzi, V. Sprio, S. Passannanti, and R. Mondelli, *Gazz. Chim. Ital.*, **98**, 907 (1968).
- F. Bohlmann, H. Suding, J. Cuatrecasas, R.M. King, and H. Robinson, *Phytochemistry*, **19**, 267 (1980).
- 13. L.F. Bjeldanes and T.A. Geissmann, Phytochemistry, 11, 327 (1972).
- F. Bohlmann, U. Fritz, R.M. King, and H. Robinson, *Phytochemistry*, 20, 743 (1981).
- 15. W. Vichnewski, E.G. Goulart, and W. Herz, *Phytochemistry*, **21**, 464 (1982).
- 16. W. Herz and N. Kumar, *Phytochemistry*, **20**, 99 (1981).
- 17. J. Gershenzon, Ph.D. dissertation, University of Texas at Austin, Austin, Texas, 1984.
- F. Melek, A. Ahmed, J. Gershenzon, and T.J. Mabry, *Phytochemistry*, 23 (in press).
- F. Bohlmann, J. Jakupovic, R.M. King, and H. Robinson, *Phytochemistry*, **19**, 863 (1980).
- G. Kreitner, J. Gershenzon, T.J. Mabry, unpublished results of work conducted at the University of Texas, Austin, 1982, and Kansas State University. (Details in reference 17.)
- T.J. Mabry, H.E. Miller, H.B. Kagan, and W. Renold, *Tetrahedron*, **22**, 1139 (1966).

Received 21 February 1984